

U.S. Application No.

International Application No.  
PCT/FR00/01318

2007 Rec'd PCT/PTO 19 NOV 2001  
09/979549  
Attorney Docket No.  
ORES10.001APC

Date: November 19, 2001

Page 1

**TRANSMITTAL LETTER TO THE UNITED STATES DESIGNATED/ELECTED OFFICE (DO/EO/US)  
CONCERNING A FILING UNDER 35 USC 371**

International Application No.: PCT/FR00/01318  
International Filing Date: May 17, 2000  
Priority Date Claimed: May 17, 1999  
Title of Invention: PROMOTER OF THIOREDOXINE TaTrxh2 IN WHEAT  
Applicant(s) for DO/EO/US: Gautier, et al.

Applicant herewith submits to the United States Designated/Elected Office (DO/EO/US) the following items and other information:

1. (X) This is a **FIRST** submission of items concerning a filing under 35 USC 371.
2. () This is a **SECOND** or **SUBSEQUENT** submission of items concerning a filing under 35 USC 371.
3. (X) This express request to begin national examination procedures (35 USC 371(f)) at any time rather than delay examination until the expiration of the applicable time limit set in 35 USC 371(b) and PCT Articles 22 and 39(1).
4. (X) A proper Demand for International Preliminary Examination was made by the 19th month from the earliest claimed priority date.
5. (X) A copy of the International Application as filed (35 USC 371(c)(2))
  - a) () is transmitted herewith (required only if not transmitted by the International Bureau).
  - b) () has been transmitted by the International Bureau.
  - c) (X) a copy of Form PCT/IB/308 is enclosed.
  - d) () is not required, as the application was filed in the United States Receiving Office (RO/US).
6. (X) A translation of the International Application into English (35 USC 371(c)(2)).
7. (X) Amendments to the claims of the International Application under PCT Article 19 (35 USC 371(c)(3))
  - a) () are transmitted herewith (required only if not transmitted by the International Bureau).
  - b) () have been transmitted by the International Bureau.
  - c) () have not been made; however, the time limit for making such amendments has NOT expired.
  - d) (X) have not been made and will not be made.
8. () A translation of the amendments to the claims under PCT Article 19 (35 USC 371(c)(3)).
9. () An oath or declaration of the inventor(s) (35 USC 371(c)(4)).
10. (X) A copy of the International Preliminary Examination Report with any annexes thereto, such as any amendments made under PCT Article 34.
11. () A translation of the annexes, such as any amendments made under PCT Article 34, to the International Preliminary Examination Report under PCT Article 36 (35 USC 371(c)(5)).

U.S. Application No.

International Application No.  
PCT/FR00/01318

09/19/2001 09:49:54  
 JC19 Rec'd PCT/PTO 19 NOV 2001  
 ORES10.001APC

Date: November 19, 2001

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12. (X) An Information Disclosure Statement under 37 CFR 1.97 and 1.98.
13. () An assignment document for recording. A separate cover sheet in compliance with 37 CFR 3.28 and 3.31 is included.
14. (X) A FIRST preliminary amendment.
15. () A substitute specification.
16. () A power of attorney and/or address letter.
17. (X) International Application as published.
18. () The present application qualifies for small entity status under 37 C.F.R. § 1.27.
19. (X) Other Items enclosed:
1. Sequence Listing in 3 pages.
  2. Sequence Listing in computer readable form (CRF)
  3. Sequence Submission Statement
  4. Notification concerning submission of Priority Document(s) PCT/IB/304.
20. (X) A return prepaid postcard.
21. (X) The following fees are submitted:

				FEES
<b>BASIC FEE</b>				\$890
<b>CLAIMS</b>	<b>NUMBER FILED</b>	<b>NUMBER EXTRA</b>	<b>RATE</b>	
Total Claims	8 - 20 =	0 ×	\$18	\$0
Independent Claims	1 - 3 =	0 ×	\$84	\$0
Multiple dependent claims(s) (if applicable)			\$280	\$0
<b>TOTAL OF ABOVE CALCULATIONS</b>				\$890
Reduction by 1/2 for filing by small entity (if applicable). Verified Small Entity statement must also be filed. (NOTE 37 CFR 1.9, 1.27, 1.28)				\$0
<b>TOTAL NATIONAL FEE</b>				\$0
<b>TOTAL FEES ENCLOSED</b>				\$890

22. (X) The fee for later submission of the signed oath or declaration set forth in 37 CFR 1.492(e) will be paid upon submission of the declaration.

U.S. Application No.

International Application No.  
PCT/FR00/01318

09/979549  
JC19 Rec'd PCT/PTO 19 NOV 2001  
Attorney Docket No.  
ORES10.001APC

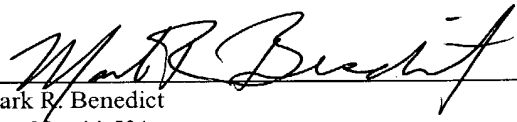
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Page 3

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23. (X) A check in the amount of \$890 to cover the above fees is enclosed.
24. () Fee for recording the enclosed assignment (37 CFR 1.21(h)). The assignment must be accompanied by an appropriate cover sheet (37 CFR 3.28, 3.31). \$40 per property.
25. (X) The Commissioner is hereby authorized to charge only those additional fees which may be required, now or in the future, to avoid abandonment of the application, or credit any overpayment to Deposit Account No. 11-1410.

**NOTE: Where an appropriate time limit under 37 CFR 1.494 or 1.495 has not been met, a petition to revive (37 CFR 1.137(a) or (b)) must be filed and granted to restore the application to pending status.**

SEND ALL CORRESPONDENCE TO:

  
Mark R. Benedict  
Reg. No. 44,531  
Customer No. 20,995

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# Knobbe Martens Olson & Bear LLP

Intellectual Property Law

09/9/9549  
JC07 Rec'd PCT/PTO 19 NOV 2001

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## CERTIFICATE OF MAILING BY "EXPRESS MAIL"

Attorney Docket No. : ORES10.001APC  
Applicant(s) : Gautier, et al.  
For : PROMOTER OF THIOREDOXINE TATRXH2 IN WHEAT  
Attorney : Mark R. Benedict  
"Express Mail"  
Mailing Label No. : EL 717638330 US  
Date of Deposit : November 20, 2001

I hereby certify that the accompanying

Transmittal;  
Demand;  
English Translation of the Application in 24 pages;  
Preliminary Amendment in 8 pages;  
Information Disclosure Statement, PTO Form 1449 with 2 references;  
Sequence listing in 3 pages and computer readable form (diskette);  
Sequence Submission Statement;  
International Preliminary Examination Report;  
International Application as published;  
Form PCT/IB/304;  
Form PCT/IB/308;  
Check for Filing Fee;  
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are being deposited with the United States Postal Service "Express Mail Post Office To Addressee" service under 37 CFR 1.10 on the date indicated above and are addressed to the Assistant Commissioner for Patents, P.O. Box 2327, Arlington, VA 22202-3514.

  
Jose Colunga

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ORES10.001APC

PATENT

## IN THE UNITED STATES PATENT AND TRADEMARK OFFICE

Applicant : Gautier, et al. )  
)  
Appl. No. : Unknown )  
)  
Filed : Herewith )  
)  
For : PROMOTER OF )  
THIOREDOXINE TaTrxh2 IN )  
WHEAT )  
)  
Examiner : Unknown )

SEQUENCE SUBMISSION STATEMENT

Assistant Commissioner for Patents  
Washington, D.C. 20231

Dear Sir:

A copy of the Sequence Listing in computer readable form as required by 37 C.F.R. § 1.821(e) is submitted herewith.

As required by 37 C.F.R. § 1.821(f), I hereby declare that the information recorded on the enclosed disk is identical to the printed Sequence Listing in the application filed herewith.

Please charge any additional fees, including any fees for additional extension of time, or credit overpayment to Deposit Account No. 11-1410.

Respectfully submitted,

KNOBBE, MARTENS, OLSON &amp; BEAR, LLP

Dated: 11/19/01By: Mark R. Benedict

Mark R. Benedict  
Registration No. 44,531  
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620 Newport Center Drive  
Sixteenth Floor  
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ORES10.001APC

PATENT

## IN THE UNITED STATES PATENT AND TRADEMARK OFFICE

Applicant	:	Gautier, et al.	)	Group Art Unit: Unknown
			)	
Appl. No.	:	Unknown	)	
			)	
Filed	:	Herewith	)	
			)	
For	:	PROMOTER OF	)	
		THIOREDOXINE TaTrxh2 IN	)	
		WHEAT	)	
			)	
Examiner	:	Unknown	)	

PRELIMINARY AMENDMENT

Assistant Commissioner for Patents  
Washington, D.C. 20231

Dear Sir:

Prior to examination on the merits, please amend the above-captioned patent application as follows:

IN THE SPECIFICATION:

On page 1, line 2, following the TITLE OF THE INVENTION, please insert the following paragraph:

CROSS REFERENCE TO RELATED APPLICATIONS

This application is a National Phase application under 35 U.S.C. § 371 of PCT application PCT/FR00/01318, filed May 17, 2000 and published in French, which claims priority to French application FR99/06231, filed May 17, 1999. The French application is incorporated herein by reference.

On page 1, line 2, before the paragraph beginning "The invention relates...", please insert the following heading:

FIELD OF THE INVENTION

009794-01 13 62 13

## BACKGROUND OF THE INVENTION

## DETAILED DESCRIPTION OF THE INVENTION

IN THE ABSTRACT:

IN THE CLAIMS:

**Please amend the claims as follows:**

- 2-





[illegible]

Respectfully submitted,

Dated:

By:

-4-

0000000000 0000000000  
Appl. No. : Unknown  
Filed : Herewith

VERSION WITH MARKINGS TO SHOW CHANGES MADE

Insertions are underlined and deletions are in **[bold and brackets]**.

IN THE SPECIFICATION:

On page 1, line 2, following the TITLE OF THE INVENTION, please insert the following paragraph:

CROSS REFERENCE TO RELATED APPLICATIONS

This application is a National Phase application under 35 U.S.C. § 371 of PCT application PCT/FR00/01318, filed May 17, 2000 and published in French, which claims priority to French application FR99/06231, filed May 17, 1999. The French application is incorporated herein by reference.

On page 1, line 2, before the paragraph beginning "The invention relates...", please insert the following heading:

FIELD OF THE INVENTION

On page 1, line 3, before the paragraph beginning "Thioredoxins are low...", please insert the following heading:

BACKGROUND OF THE INVENTION

On page 1, line 12, before the paragraph beginning "In plants...", please insert the following heading:

DETAILED DESCRIPTION OF THE INVENTION

On page 23, line 1, please omit the heading "Claims" and insert the following heading:  
WHAT IS CLAIMED IS:

IN THE ABSTRACT:

Please insert the abstract attached hereto, following the page entitled VERSION WITH MARKINGS TO SHOW CHANGES MADE, as page 25 of the specification.

090795Z . 06130

**Please cancel Claim 8.**

**Please amend the claims as follows:**

1. A p[P]romoter, comprising[characterized in that it consists of] a nucleic acid fragment comprising at least one specific functional domain of the promoter of the *TaTrxh2* gene.
2. The p[P]romoter according to Claim 1, wherein[characterized in that] said nucleic acid fragment is chosen from the group consisting of:
  - the nucleic acid fragment, the sequence of which extends from position -1 to position -1111 relative to the ATG codon of the *TaTrxh2* gene;
  - the nucleic acid fragment, the sequence of which extends from position -1 to position -83 relative to the ATG codon of the *TaTrxh2* gene;
  - the nucleic acid fragment, the sequence of which extends from position -451 to position -591 relative to the ATG codon of the *TaTrxh2* gene;
  - the nucleic acid fragment, the sequence of which extends from position -591 to position -1111 relative to the ATG codon of the *TaTrxh2* gene;
  - the nucleic acid fragment, the sequence of which extends from position -228 to position -451 relative to the ATG codon of the *TaTrxh2* gene;
  - the nucleic acid fragment, the sequence of which extends from position -451 to position -591 relative to the ATG codon of the *TaTrxh2* gene;
  - the nucleic acid fragment, the sequence of which extends from position -83 to position -228 relative to the ATG codon of the *TaTrxh2* gene;
  - the nucleic acid fragment, the sequence of which is that of the first intron of the *TaTrxh2* gene.
3. An e[E]xpression cassette, comprising[characterized in that it comprises] a promoter according to [either of ]Claim[s] 1[ and 2].
4. A r[R]ecombinant vector, comprising[characterized in that it comprises] at least one promoter according to [either of ]Claim[s] 1[ and 2].
5. A p[P]lant cell transformed with at least one promoter according to [either of]Claim[s] 1[ and 2].

**06-07-2018**

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## PROMOTER OF THIOREDOXINE TaTrxh2 IN WHEAT

The invention relates to the cloning and to the characterization of a wheat thioredoxin promoter.

Thioredoxins are low molecular weight proteins which have been demonstrated in a large number of organisms, in which they catalyze various redox reactions involving dithiolsulphhydryl exchanges. Their catalytic site comprises the conserved sequence: -Trp-Cys-Gly/Pro/Ala-Pro-Cys-. Thioredoxins in oxidized form comprise a disulphide bridge, the reduction of which to -SH groups, by reduced ferredoxin or by NADPH, is catalyzed via a specific system.

In plants, 3 types of thioredoxin have been demonstrated: the first 2 (thioredoxins m and f) are ferredoxin-dependant thioredoxins located in the chloroplasts, where they are involved in the regulation of photosynthesis. A third type, named thioredoxin h, has been demonstrated in the cytosol. Thioredoxin h is part of an NADP-dependent thioredoxin system (NTS), in which it is associated with NADPH and with an enzyme named NADP-thioredoxin reductase (NTR).

Initially, 2 thioredoxin h were extracted and partially purified from wheat grain (VOGT and FOLLMANN, Biochem. Biophys. Acta 873, 415-418, 1986). Recently, the inventors' team has isolated and characterized 2 cDNA clones encoding a soft wheat thioredoxin h (TaTrxhl) and a hard wheat thioredoxin h (TdTrxhl) (GAUTIER *et al.*, Eur. J. Biochem. 252, 314-324, 1998). The primary structures deduced from the cDNA clones of the thioredoxins h TaTrxhl and TdTrxhl are very conserved (96% identity between them). They have an N-terminal extension which is very rich in Ala residues, the analysis of which reveals a putative transmembrane domain of 20 residues. They show strong homologies with cereal thioredoxin h (70-80%) and dicotyledon thioredoxin h (60%).



demonstrated regions involved in the spatial and temporal regulation of this promoter.

The sequence of the *TaTrxh2* gene and of the region in 5' comprising the promoter are represented in the attached sequence listing under the number SEQ ID NO: 1.

A subject of the present invention is a promoter consisting of a nucleic acid fragment comprising at least one specific functional domain of the promoter of the *TaTrxh2* gene.

The term "promoter" is intended to mean a double-stranded DNA sequence comprising at least the sequences required for initiating the transcription of a gene, optionally combined with sequences for regulating, in *cis*, said transcription; the expression "specific functional domain of a promoter" is intended to mean a sequence of said promoter comprising one or more DNA units involved in transcription initiation, or a double-stranded DNA sequence constituting a regulatory domain comprising one or more of the DNA units involved in the control, in *cis*, of transcription by said promoter.

Promoters in accordance with the invention may, in particular, comprise:

a) the nucleic acid fragment which is represented in the attached sequence listing by the sequence SEQ ID NO: 2, and also on Figure 1, and which corresponds to the 5' non-coding region of the *TaTrxh2* gene, extending from position -1 to position -1111 relative to the ATG initiation codon, or portions of said fragment, in particular:

\* the nucleic acid fragment, the sequence of which extends from position -1 to position -83 relative to the ATG codon of the *TaTrxh2* gene; this fragment comprises the sequences involved in transcription initiation and required for the basic activity of the promoter;

\* nucleic acid fragments comprising functional domains involved in regulating the transcription of the *TaTrxh2* gene, and in particular involved in its tissue specificity and/or in its expression at various stages of the development of the plant; it is in particular:

- the nucleic acid fragment, the sequence of which extends from position -591 to position -1111 relative to the ATG codon of the *TaTrxh2* gene; this fragment comprises a regulatory domain involved in inhibiting the expression of the *TaTrxh2* gene in the scutellum epithelium;

- the nucleic acid fragment, the sequence of which extends from position -228 to position -451 relative to the ATG codon of the *TaTrxh2* gene; this fragment comprises a regulatory domain involved in inducing the expression of the *TaTrxh2* gene at the beginning of maturation of the grain;

- the nucleic acid fragment, the sequence of which extends from position -451 to -591 relative to the ATG codon of the *TaTrxh2* gene; this fragment comprises a regulatory domain involved in inducing the expression of the *TaTrxh2* gene in the scutellum epithelium;

- the nucleic acid fragment, the sequence of which extends from position -83 to position -228 relative to the ATG codon of the *TaTrxh2* gene; this fragment comprises sequences involved in inducing the expression in the starch albumen.

b) the nucleic acid fragment constituting the first intron (positions 1232-2203 on the sequence SEQ ID NO: 1) of the *TaTrxh2* gene; this fragment may comprise a regulatory domain of the amplifier type, which increases the level of expression of the *TaTrxh2* gene.

Those skilled in the art may, using the fragments comprising at least one functional domain of the promoter of the *TaTrxh2* gene, specified above, identify more precisely the limits of these functional



domains, and also the DNA units involved in the function of each of them, using techniques which are, in themselves, known, for example using the DNA footprinting technique, incubating these fragments with nuclear  
5 extracts of cells from the albumen of the grain, and also with nuclear extracts of cells in which the promoter of the *TaTrxh2* gene is inactive.

The invention in particular encompasses any promoter which may be obtained from a nucleic acid  
10 fragment comprising at least one functional domain of the promoter of the *TaTrxh2* gene, using conventional genetic engineering techniques, in particular by mutagenesis and/or genetic recombination. It is thus possible to produce artificial promoters which have the desired level  
15 of activity and degree of specificity.

It is thus possible, for example, to inactivate one or more of the functional regulatory domains located in the 5' non-coding region of the *TaTrxh2* gene, for example by deleting at least one  
20 nucleotide or a sequence of nucleotides of the DNA units involved in the function of the domain(s) in question. It is also possible to combine the nucleic acid molecules comprising functional domains of the promoter of the *TaTrxh2* gene with one another, and/or with functional  
25 domains originating from promoters other than that of the *TaTrxh2* gene.

According to a preferred embodiment of a promoter in accordance with the invention, it comprises at least the sequences of the promoter of *TaTrxh2* which  
30 allow specific expression in the grain, in particular in the starchy albumen.

The invention also encompasses:

- the expression cassettes comprising, besides a promoter in accordance with the invention, a gene of  
35 interest placed under the transcriptional control of said

promoter, or a site which allows the insertion of said gene of interest;

- the recombinant vectors resulting from the insertion of a promoter or of an expression cassette in accordance with the invention into a host vector.

The promoters in accordance with the present invention may be used to control the expression of a gene of interest in plant cells, in particular monocotyledon cells.

Said gene of interest may, for example, be either the *TaTrxh2* gene, placed under the control of an artificial promoter, as defined above, derived from the *TaTrxh2* promoter, or a heterologous gene encoding a thioredoxin other than *TaTrxh2*, or any other protein of interest.

It is possible, for example, to introduce a gene of interest under the control of the promoter of the *TaTrxh2* gene, or of an artificial promoter constructed from the regulatory elements thereof which confer the specificity of expression in the albumen cells, in order to express said gene of interest only in the grain albumen cells. It is also possible to selectively delete the sequences of the promoter of the *TaTrxh2* gene which are responsible for the specificity of expression, in order to construct an artificial promoter which makes it possible to ensure ubiquitous expression of a thioredoxin h, or of another protein of interest.

A subject of the invention is also plant cells and transgenic plants, in particular monocotyledons, and especially cereals, transformed with at least one nucleic acid molecule comprising a promoter in accordance with the invention.

The inventors have thus obtained transgenic rice plants in which a heterologous gene has been placed under the transcriptional control of the promoter of the

*TaTrxh2* gene and have observed, in these plants, specific expression in the grain albumen cells.

The transformed cells and the transgenic plants in accordance with the invention can also be used  
 5 as models to study and/or modify the expression of various genes in the grain albumen cells.

The present invention will be more clearly understood using the further description which follows, which refers to nonlimiting examples illustrating the  
 10 cloning and characterization of the *TaTrxh2* gene and of its promoter.

#### **EXAMPLE 1: ISOLATION AND CHARACTERIZATION OF THE *TaTrxh2* GENE**

##### **1.- Screening a wheat genomic DNA library**

15 A genomic DNA library was prepared from the DNA extracted from leaves of soft wheat (*Triticum aestivum*) of the variety Andain. After partial digestion of the genomic DNA with *MboI*, the fragments with a mean size of 15 kb were cloned at the *BamHI* site of the EMBL3  
 20 SP6/T7 phage, which was propagated in the host bacterium K802 -K 802 (*galK2*, *galT22*, *HsdR2*, (*r<sub>k</sub>*<sup>-</sup>, *m<sub>k</sub>*<sup>+</sup>), *mcrA*<sup>-</sup>, *mcrB*<sup>-</sup>, *metB1*, *mrr*<sup>+</sup>, *supE44*).

6 × 10<sup>6</sup> clones from the genomic DNA library were plated out and screened with a 669 bp probe (TRX)  
 25 containing all of the sequence encoding the soft wheat thioredoxin h *TaTrxh1* (GAUTIER et al., 1998, publication mentioned above), and the positive clones were then screened by PCR (polymerase chain reaction) using a pair of primers (THP2 and THM2) derived from the same  
 30 sequence.

One of the clones selected (λ4), which contains an approximately 10 kb wheat genomic DNA fragment, was digested with *PstI*, releasing two fragments, one of 1.5 kb and the other of 3.8 kb; both  
 35 recognized by the TRX probe. These two fragments were

cloned into the pLITMUS 29 vector (BIOLAB) at the *Pst*I restriction site. The two clones obtained are named CTRX3 and CTRX4. The CTRX3 clone corresponds to the 1.5 kb fragment and the CTRX4 clone to the 3.8 kb fragment.

5           Analysis of the nucleotide sequences of the CTRX4 and CTRX3 clones shows that each contains part of the same gene encoding a wheat thioredoxin h, which is truncated during digestion with *Pst*I.

10           Based on the nucleotide sequences of these two clones, the inventors chose two primers (THP8 and THM8) making it possible to amplify a thioredoxin h gene over a length of approximately 2.6 kb. The PCR was carried out on the undigested DNA of the  $\lambda$ 4 clone, and a fragment of the expected size was cloned into the pGEM-T vector  
15 (PROMEGA). The clone obtained contained the *TaTrxh2* gene encoding a soft wheat thioredoxin h.

          It comprises a 1111 bp promoter region, a 1447 bp coding region and a 131 bp 3' non-coding region.

20           The coding region of the *TaTrxh2* gene comprises three exons 120, 123 and 135 bp, separated by two introns of 972 bp and 93 bp. The first exon encodes a 40 amino acid polypeptide, the second exon encodes a 41 amino acid polypeptide containing the active site, and the third exon encodes a 45 amino acid polypeptide.

25           The nucleotide sequence of the *TaTrxh2* gene encodes a soft wheat thioredoxin h named *TaTrxh2*, which has 126 amino acids, a calculated molecular mass of 13435 Da and a calculated pI of 5.0.

30           Comparison of the sequences of the translation products of the *TaTrxh2* gene and of the genes *TaTrxh1* previously described by GAUTIER *et al.* (1998, publication mentioned above) and *TdTrxh1* (hard wheat thioredoxin h) shows that they are very conserved. Specifically, the peptide sequence of *TaTrxh2* exhibits 97% similarity and  
35 94% identity with that of *TaTrxh1* and 95% similarity and 90% identity with that of *TdTrxh1*.

The N-terminal domain of TaTrxh2 is shorter than that of TaTrxh1 and TdTrxh1. The primary structure of TaTrxh2 does not contain a signal peptide, suggesting that the protein is located in the cytoplasm. However, it has an N-terminal extension already demonstrated in the primary structure of TaTrxh1 and TdTrxh1, which may correspond to a transmembrane domain. Analysis of the N-terminal extension of TaTrxh2 with the RAO ARGOS program (PC/gene, RAO *et al.*, Biochem. Biophys. Acta 869, 197-214, 1986) reveals a putative transmembrane domain between residues 2 and 19. The active site, made up of the following 5 amino acids: WCGPC, is conserved between the three wheat thioredoxin h TaTrxh2, TaTrxh1 and TdTrxh1.

The sizes of the introns are different from those of the introns of the wheat thioredoxin h genes previously demonstrated by ROBERT (1994), indicating that the TaTrxh2 gene is different from those genes. The introns of the TaTrxh2 gene are of the 0 type and are limited in 5' by the sequence GTA and in 3' by the sequence CAG, which correspond to consensus sequences of intron-exon limits.

The 3' non-coding region of the TaTrxh2 gene has the polyadenylation signal AATAAA common to genes transcribed by RNA polymerase II.

#### **EXAMPLE 2: STRUCTURAL ANALYSIS OF THE PROMOTER OF THE TaTrxh2 GENE**

The promoter of the TaTrxh2 gene was analysed in order to search for putative regulatory units likely to be involved in controlling expression, and was in particular compared to that of the thioredoxin h genes of *C. reinhardtii* (STEIN *et al.*, Plant Mol. Biol. 28, 487-503, 1995), of tobacco (BRUGIDOU *et al.*, Mol. Gen. Genet. 238, 285-293, 1993) and of rice (ISHIWATARI *et al.*, 1995), of the thioredoxin m gene of *C. reinhardtii* (STEIN *et al.*, 1995), and of the murine (MATSUI *et al.*, Gene

152, 165-171, 1995) and human (TONISSEN *et al.*, Gene 102, 221-228, 1992; KAGHAD *et al.*, Gene 140, 6643-6653, 1994) thioredoxins.

5 The sequence of the promoter of the *TaTrxh2* gene is represented on Figure 1.

The transcription initiation site (represented on Figure 1 in bold and underlined with a double line) is an adenine located at -65 bp from the ATG.

10 The promoter of the *TaTrxh2* gene contains no consensus sequence corresponding to a TATA box or to a CAAT box at the expected positions for genes transcribed by RNA polymerase II.

15 On the other hand, it contains a TATA-like box (AATTTAT, underlined with a double line on Figure 1) at -105 bp from the ATG.

20 It also contains a GC box (GGGCCGGG, underlined with a dotted line on Figure 1) located at -84 bp from the ATG of the *TaTrxh2* gene. GC boxes are recognized by transcription factors of the Sp1 type (DYNAN *et al.*, Nature 316, 774-778, 1985) and are involved in the constitutive expression of genes. GC boxes are present in all the known promoters of thioredoxin genes.

25 A sequence rich in adenine, interrupted by a G residue (AAAAAAGAAAAA, in bold characters underlined with a single line on Figure 1), is located at -227 bp from the ATG of the *TaTrxh2* gene; sequences of this type have also been identified previously in the promoters of the tobacco and rice thioredoxin h genes.

30 *bHLH* sequences (CANNTG), recognized by transcription factors of the helix/loop/helix family, are located at -206 bp and -411 bp from the ATG of the *TaTrxh2* gene; they are represented on Figure 1 in lower case letters.

35 *bzip* sequences (ACGT, underlined with a single line on Figure 1), recognized by transcription factors of

the leucine zipper family (bZIP), are located at -251 bp and -184 bp from the ATG of the *TaTrxh2* gene. The bZIP proteins have been described in the activation of expression of genes encoding grain storage proteins. ACGT units have also been described in ABRE (ABA-responsive element) consensus sequences of the promoters of genes, the expression of which is regulated by abscissic acid (ABA) (MUNDY *et al.*, Proc. Natl. Acad. Sci. USA, 87, 1406-1410, 1990).

Two pyrimidine boxes (CCTTTCTCT and TCTTTCTTC, boxed on Figure 1) are, respectively, located at -553 bp and -541 bp from the ATG of the *TaTrxh2* gene. Pyrimidine boxes (CCTTTT) are involved in the regulation of expression by giberellic acid, generally in combination with GARE (GA-responsive element) sequences (TAACAAA) (HUANG *et al.*, Plant Mol. Biol. 14, 115-121, 1990), and O2S (opaque-2-binding sequence) or I box (TATCCAT) sequences (GUBLER *et al.*, Plant Cell 4, 1435-1441, 1992; LANAHAHAN *et al.*, Plant Cell, 4, 203-211, 1992), with which they organize into a complex named GARC (GA-responsive complex) (BETHKE *et al.*, Bot. 48, 1337-1356, 1997).

No GARE or O2S sequence was revealed in the 1111 bp upstream of the ATG of the *TaTrxh2* gene.

A TGTGTGAGCA unit (in bold characters and underlined with a dotted line on Figure 1) is located at -403 bp from the ATG of the *TaTrxh2* gene. This unit differs only by the presence of an additional G residue, from the GCN4-like consensus sequence (TGTGTGACA) of the "albumen box" involved in the albumen-specific expression of wheat glutenin genes (HAMMOND-KOSACK *et al.*, EMBO J. 12, 545-554, 1993). However, the other unit of the albumen box, named EM (TGTAAGAGT), and the presence of which is also required for albumen-specific expression (ALBANI *et al.*, Plant Cell 9, 171-184, 1997), was not revealed in the 1111 bp upstream of the ATG of the *TaTrxh2* gene.

Trimeric diades CAA and TTG (in italics on Figure 1) separated by 10 bases are present, respectively, at -107 bp and -97 bp from the ATG of the *TaTrxh2* gene. These units are associated with specific expression in the aleurone layer (THOMAS et al., Plant Cell 2, 1171-1180, 1990).

**EXAMPLE 3: FUNCTIONAL ANALYSIS OF THE PROMOTER OF THE *TaTrxh2* GENE.**

The 1111 bp sequence in 5' of the ATG of the *TaTrxh2* gene, or various fragments of this sequence, were cloned upstream of the coding sequence of the *gus* reporter gene in the pSPORT1-GUS vector. The pSPORT1-GUS vector (DIGEON, 1997) contains the coding sequence of the *gus* (*E. coli*  $\beta$ -glucuronidase) gene and the *nos-ter* terminator of the nopaline synthase gene, inserted at the EcoRI-HindIII site of the pSPORT1 vector (GIBCO BRL).

The constructs prepared are as follows:

- P1: this construct comprises all of the 1111 bp sequence upstream of the ATG of the *TaTrxh2* gene;
- P2: this construct comprises the 589 bp sequence upstream of the ATG of the *TaTrxh2* gene;
- P3: this construct comprises the 481 bp sequence upstream of the ATG of the *TaTrxh2* gene;
- P4: this construct comprises the 228 bp sequence upstream of the ATG of the *TaTrxh2* gene;
- P5: this construct comprises the 83 bp sequence upstream of the ATG of the *TaTrxh2* gene;

The limits of the regions of the promoter of the *TaTrxh2* gene which were used in the constructs are indicated on Figure 1.

As a positive control, the pUGC1 vector (CHAÏR et al., 1996), which allows constitutive and ubiquitous expression of the *gus* gene under the control of the promoter, of the first exon and of the first intron of the gene encoding maize ubiquitin was used.



The pSPORT1-GUS vector was used as a negative control.

These various constructs were transferred, by bombardment according to the protocol described by FAUQUET *et al.* (Proc. Third. Int. Rice Genet. Symp., Ed. G.S. Khush, 153-165, 1996), into young embryogenic calluses of rice (var. japonica IRAT 349) which derive from the proliferation of the scutellum of the mature embryo. All the constructs tested were cotransferred with the pILTAB227 vector (FAUQUET *et al.*, 1997), which confers hygromycin resistance and which allows selection of the transformed cells.

A mixture of the vector carrying the construct to be tested and of the pILTAB227 vector (vector to be tested/pILTAB227 molar ratio = 4/1) is used to coat gold microparticles, in a proportion of 5 µg of total DNA (3 µg of DNA to be tested + 2 µg of pILTAB227) at a concentration of 1 µg/µl, per 3 mg of a mixture, in equal amounts, of gold microparticles having diameters of 1.0 µm and 1.6 µm, in suspension in 50 µl of distilled water.

The bombardment is carried out using a PDS-1000/He particle gun (PARTICLE DELIVERY SYSTEM, BIORAD).

The bombarded embryogenic calluses are then screened on a selection medium containing hygromycin. The hygromycin-resistant calluses are selected and placed on hygromycin-free regeneration medium. The regenerated plants (F0 generation) are then transferred into pots and, after acclimatization in a phytotron, are cultivated under glass.

The expression of the *gus* gene was sought in the vegetative organs and in the grains of the rice plants of the T0 and T1 generations. Only the plants which were fertile and had a normal phenotype were selected for analysis. Integration of the transgene into

the plants analysed was verified by PCR and Southern transfer.

The  $\beta$ -glucuronidase activity was detected using a histochemical test, detecting the blue coloration resulting from hydrolysis of 5-bromo-4-chloro-3-indolyl  $\beta$ -D-glucuronic acid (X-GLU), and was quantified using a fluorometric test, measuring the 4-methylumbelliferone (MU) formed from 4-methylumbelliferyl  $\beta$ -glucuronic acid, according to the protocol described by JEFFERSON *et al.* (Plant Mol. Biol. Report 5, 387-405, 1987).

### **1. Expression of the *gus* gene in the vegetative organs**

The analysis was carried out on the roots, the culms and the leaves.

In the case of the nontransformed rice plants, or of those transformed with the pSPORT1-GUS vector, the histochemical analysis reveals no GUS activity, and the fluorimetric measurements reveal only very low, or even zero, activity.

In the case of the rice plants transformed with the pUGC1 vector, high GUS activity (greater than 500 pmol MU/min/mg of protein) is observed in all the vegetative organs tested.

In the case of the rice plants transformed with the P1, P2, P3, P4 or P5 constructs, no coloration is observed in the vegetative organs incubated in the presence of X-GLU, and the GUS activity measured by fluorometry is not significantly different from those measured for the plants transformed with the pSPORT1-GUS vector or the nontransformed plants. These results show that the promoter of the *TaTrxh2* gene does not allow expression of the *gus* gene in the vegetative organs.

## 2. Expression of the *gus* gene in the grains

## Histochemical analysis

*In whole grains*

The rice grains, taken at 35 DAF (days after  
5 fertilization) were cut in the longitudinal direction and  
then incubated in the presence of X-GLU.

No coloration is detected in the rice grains transformed with the pSPORT1-GUS vector.

An intense coloration of the entire grain is, on the contrary, detected for the rice grains transformed with the pUGC1 vector.

In the case of the rice grains transformed with the P1, P2, P3 or P4 constructs, a blue coloration is detected in the albumen of the grains, but not in the embryo (cotyledonary axis and scutellum), the envelopes or the spikelets. In the albumen, this coloration appears in particular at the periphery of the embryo, above the scutellum epithelium, and in a median zone of the albumen over the entire length of the grain.

20                    This coloration is less intense and appears  
less rapidly than that observed in the rice grains  
transformed with pUGC1. The intensity of the coloration  
appears to vary depending on the construct (P1, P2, P3 or  
P4) concerned. The strongest intensity is observed in the  
25 rice grains transformed with the P2 construct and the  
weakest intensity is observed in those transformed with  
the P4 construct. These results are confirmed by the  
analysis of the T1 grains, in which the coloration  
appears more rapidly than in the T0 grains and is more  
30 intense.

In the case of the rice grains transformed with the P5 construct, no coloration is detected, either in the embryo, in the albumen or in the envelopes.

In the various tissues of the grain

In order to accurately determine the location of the expression of the *gus* gene under the control of the promoter of the *TaTrxh2* gene, histological sections were prepared and observed using a photon microscope.

The observations show that, for the P1, P2, P3 or P4 constructs, the labelling is located in a small number of cells of the starchy albumen, located at the periphery of the embryo and in the central zone of the starchy albumen. The cells of the embryo, of the aleurone layer and of the envelopes are not labelled. For the P5 construct, no labelling is visible.

### Fluorimetric analysis

The GUS activity was measured, firstly, on the  
15 embryos and, secondly, on the albumen of the rice grains.

The GUS activity is zero or very low in the rice grains transformed with the pSPORT1-GUS vector, as in the nontransformed rice grains. On the other hand, it is very high ( $>500$  pmol/MU/min/mg of protein) in the embryo and the albumen of the rice grains transformed with the pUGC1 vector.

The GUS activity measured in the embryos of the rice grains transformed with the P1, P2 P3, P4 or P5 constructs is not significantly different from that measured in the nontransformed rice grains or rice grains transformed with the pSPORT1-GUS vector.

On the other hand, the activity measured in the albumen of the rice grains transformed with the P1, P2, P3 or P4 constructs is 25 to 40 times higher than that measured in the albumen of the nontransformed rice grains or the rice grains transformed with the pSPORT1-GUS vector. It ranges from 40 pmol/MU/min/mg of protein, for the rice grains transformed with the P2 construct, to 25 pmol/MU/min/mg of protein, for those transformed with the P4 construct. For the P5 construct, no GUS activity is detected in the grains.

These results show that the region (-1111 bp to -83 pb) of the promoter of the *TaTrxh2* gene allows expression of the *gus* gene only in the cells of the starchy albumen, and that only the deletion which leaves  
 5 only 83 bp upstream of the ATG suppressed the sequences responsible for the spatial expression, some of which are probably located in the region of the promoter which is between -228 bp and -83 pb.

The GCN4-like unit identified during the  
 10 structural analysis of the promoter of the *TaTrxh2* gene is therefore apparently not the only one responsible for the tissue specificity of the expression; specifically, despite being deleted in the P3 and P4 constructs, the expression of the *gus* gene remains specific for the  
 15 albumen of the grain.

Two sequences: AACAAATCC and AACAAAGTG (represented in bold characters on Figure 1), are present, at -51 bp and -381 bp, respectively, relative to the ATG of the *TaTrxh2* gene. These sequences exhibit similarity  
 20 with AACA units (AACAAACTCTATC) recently demonstrated in the promoters of 6 genes encoding rice glutelins, and involved in the albumen-specific expression of these genes.

#### 25 **EXAMPLE 4: EVOLUTION OF THE EXPRESSION OF THE GUS GENE DURING THE DEVELOPMENT OF THE GRAINS OF THE TRANSGENIC RICE PLANTS**

The expression of the *gus* gene was monitored during the maturation and germination of rice grains transformed with the P1, P2, P3, P4 or P5 constructs.

##### 30 **1. During maturation**

Three stages were analysed: 10 DAF, 25 DAF and 35 DAF. The expression of the *gus* gene was evaluated either by histochemical location of the GUS activity or by detection of the transcripts by Northern transfer.

GUS activity

The histochemical analysis shows that, for the three stages of maturation studied, 10, 25, and 35 DAF, GUS activity is always detected in the starchy albumen of the rice grains transformed with the P1, P2, P3 or P4 constructs. On the other hand, for the P5 construct, no GUS activity is detected.

At 10 DAF, the GUS activity is detected in the starchy albumen, at the periphery of the embryo. At 25 DAF, the GUS activity progresses towards the median zone of the starchy albumen. At 35 DAF, the GUS activity is detected over the entire surface of the starchy albumen.

The intensity of the coloration varies with the nature of the construct and the stage of maturation, in particular in the case of the P4 construct, for which the coloration is very difficult to detect at the start of maturation.

These results are confirmed by the more detailed observations in each tissue of the grain, which show that:

- At 10 DAF: for the P1, P2 or P3 constructs, the GUS activity is very high in the cells of the starchy albumen at the periphery of the embryo, and it is not detected in the cells of the median zone of the starchy albumen. For the P4 construct, the GUS activity in the cells of the starchy albumen is very low, or even undetectable. In addition, in the rice grains transformed with the P2 construct, GUS activity is also detected in the cells of the scutellum epithelium.

- At 25 DAF: for the P1, P2 or P3 constructs, the GUS activity decreases in the cells of the starchy albumen at the periphery of the embryo and increases in those of the central zone of the starchy albumen; for the rice grains transformed with the P2 construct, GUS activity is no longer detected in the cells of the

scutellum epithelium. In the rice grains transformed with the P4 construct, the GUS activity increases in the cells of the starchy albumen which are located at the periphery of the embryo and in the central zone.

5                   - At 35 DAF: the GUS activity is much lower than at 25 DAF in all the cells of the starchy albumen of the rice grains transformed with P1, P2 or P3; in the rice grains transformed with the P4 construct, the GUS activity is detected in the cells of the median zone of  
10 the starchy albumen.

For the P5 construct, no GUS activity is detected, whatever the stage of maturation or the tissue of the grain analysed.

Whatever the construct used, no GUS activity  
15 is detected, during maturation, in the cells of the embryonic axis, of the aleurone layer or of the envelopes of the grains.

#### Detection of the *gus* gene transcripts

The presence of the *gus* gene transcripts was  
20 sought in the total RNAs extracted, at the various stages of maturation, from the rice grains transformed with the P1, P2, P3, P4 or P5 constructs. The detection was carried out by Northern transfer, using the P3+GUS probe. This 2.6 kb probe comprises the 481 bp sequence upstream  
25 of the ATG of the *TaTrxh2* gene, the coding sequence of the *gus* gene and the terminator of the *nos* gene.

For each of the P1, P2, P3, P4 or P5 constructs, this probe makes it possible to detect the presence of transcripts which have an expected size of  
30 between 1.9 and 2.4 kb, depending on the construct.

The presence of these transcripts varies depending on the construct used for the transformation and on the stage of maturation.

For the rice plants transformed with the P1  
35 construct, the transcripts are detected at the 3 stages of maturation, with a maximum at mid-maturation.

For the rice plants transformed with the P2 construct, the transcripts are detected at the start of maturation.

5 For the rice plants transformed with the P3 construct, the transcripts are detected at the start and at the end of maturation of the grain.

For the rice plants transformed with the P4 construct, the transcripts are detected at mid-maturation and at the end of maturation of the grain.

10 For the rice plants transformed with the P5 construct, no *gus* gene transcript is detected, whatever the stage of maturation analysed.

## **2. During germination**

15 For the study of the expression of the *gus* gene during germination, the GUS activity was analysed by histochemistry in rice grains transformed with the P1, P2, P3, P4 or P5 constructs.

For each construct, 10 grains were left to germinate in the dark and taken at various times after soaking: 0, 12, 24, 48 and 72 hours.

20 The GUS activity is detected in the starchy albumen of the rice grains transformed with the P1, P2, P3 or P4 constructs, whatever the stage of germination. On the other hand, for the rice grains transformed with the P5 construct, no GUS activity is detected.

30 The study of the accumulation of the *gus* gene transcripts in the grains transformed with the P1, P2, P3 or P4 constructs shows that these transcripts are not accumulated during germination, regardless of the construct.

These results indicate that the promoter (1111 bp upstream of the ATG) of the *TaTrxh2* gene does not allow expression of the *gus* gene during germination. The GUS activity detected in the germinated grains is 35 certainly a residual activity due to the very great stability of  $\beta$ -glucoronidase in the grain.



### Conclusion

The analysis of the expression of the *gus* gene under the control of the promoter of the *TaTrxh2* gene, during the development of the rice grains transformed with the P1, P2, P3, P4 or P5 constructs demonstrates an effect of the deletions of the promoter of the *TaTrxh2* gene on the temporal and spatial expression of the *gus* gene in the grains of the rice plants transformed.

The P1, P2 and P3 constructs allow earlier expression of the *gus* gene than the P4 construct, during maturation. The P5 construct does not allow expression of the *gus* gene since no transcript is detected. In fact, *gus* gene transcripts are detected at 10 DAF for the 3 constructs P1, P2 and P3, and only 25 DAF for the P4 construct. This suggests that the differences in level of expression previously noted between the P2 and P4 constructs are probably the result of a delay in the expression of the *gus* gene under the control of the P4 promoter, rather than the result of a lower level of expression. The region of the promoter of the *TaTrxh2* gene which is between -1111 bp and -228 bp definitely contains regulatory sequences which allow expression of the *gus* gene in the first stages of maturation.

With regard to the spatial expression, the region of the promoter of the *TaTrxh2* gene which is between -1111 bp and -591 bp probably contains a sequence which inhibits the expression of the gene in the scutellum epithelium. In fact, when it is deleted (P2 construct), expression of the *gus* gene is observed in this tissue. Conversely, the region between -591 bp and -451 bp is thought to contain a sequence which activates the expression in the scutellum epithelium since, when it is deleted (P3 construct), there is no longer an expression of the *gus* gene in this tissue.

The results show that, during the maturation of the rice grains, the promoter of the *TaTrxh2* gene

5

1) Promoter, characterized in that it consists of a nucleic acid fragment comprising at least one specific functional domain of the promoter of the *TaTrxh2* gene.

10       - the nucleic acid fragment, the sequence of  
which extends from position -1 to position -1111 relative  
to the ATG codon of the *TaTrxh2* gene;

15           - the nucleic acid fragment, the sequence of  
which extends from position -451 to position -591  
relative to the ATG codon of the *TaTrxh2* gene;

- the nucleic acid fragment, the sequence of which extends from position -591 to position -1111 relative to the ATG codon of the *TaTrxh2* gene;

- the nucleic acid fragment, the sequence of which extends from position -228 to position -451 relative to the ATG codon of the *TaTrxh2* gene;

25        - the nucleic acid fragment, the sequence of  
which extends from position -451 to position -591  
relative to the ATG codon of the *TaTrxh2* gene;

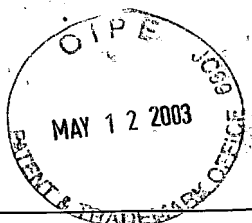
- the nucleic acid fragment, the sequence of which extends from position -83 to position -228 relative to the ATG codon of the *TaTrxh2* gene;

30                   - the nucleic acid fragment, the sequence of  
which is that of the first intron of the *TaTrxh2* gene.

3) Expression cassette, characterized in that it comprises a promoter according to either of Claims 1 and 2.

10                    8) Use of a promoter according to either of  
Claims 1 and 2, for controlling the expression of a gene  
of interest in a plant cell.

The invention relates to the cloning and characterization of a wheat thioredoxin promoter, specifically to the promoter of thioredoxin TaTrxh2 in wheat.



Attorney's Docket No. \*

ORES10.001AFC  
Filed 11-19-01**DECLARATION AND POWER OF ATTORNEY – USA PATENT APPLICATION**

As a below named inventor, I hereby declare that:

My residence, post office address and citizenship are stated below next to my name;

I believe I am the original, first and sole inventor (if only one name is listed below) or an original, first and joint inventor (if plural names are listed below) of the subject matter which is claimed and for which a patent is sought on the invention entitled **PROMOTER OF THIOREDOXINE TaTrxh2 IN WHEAT**; the specification of which is attached hereto;

I hereby state that I have reviewed and understand the contents of the above identified specification, including the claims;

I acknowledge the duty to disclose information which is material to the examination of this application in accordance with Title 37, Code of Federal Regulations, § 1.56(a);

I hereby claim foreign priority benefits under Title 35, United States Code, § 119 of any foreign application(s) for patent, design or inventor's certificate having a filing date before that of the application on which priority is claimed:

**PRIOR FOREIGN APPLICATION(S)****Priority Claimed**

No.:	<u>99/06231</u>	Country:	<u>France</u>	Date Filed:	<u>May 17, 1999</u>	<b>YES</b>
No.:	_____	Country:	_____	Date Filed:	_____	
No.:	_____	Country:	_____	Date Filed:	_____	

I hereby claim the benefit under Title 35, United States Code, § 120 of any United States application(s) listed below, and insofar as the subject matter of each of the claims of this application is not disclosed in the prior United States application in the manner provided by the first paragraph of the Title 35, United States Code § 112, I acknowledge the duty to disclose material information as defined in Title 37, Code of Federal Regulations, § 1.56(a) which occurred between the filing date of the prior application and the national or PCT international filing date of this application :

**Prior U.S.A. Application(s)**

Serial No.: \_\_\_\_\_ Filing date: \_\_\_\_\_ Status: \_\_\_\_\_

(25) **POWER OF ATTORNEY:** I hereby appoint the following attorneys and/or agents to prosecute this application and to transact all business in the Patent and Trademark Office connected therewith (if this application is assigned, I acknowledge that the appointed individuals do not represent me, and that instead they represent the assignee): Louis J. Knobbe, Registration No. 18,780; Don W. Martens, Registration No. 21,107; Gordon H. Olson, Registration No. 20,319; James B. Bear, Registration No. 25,221; Darell L. Olson, Registration No. 28,247; William B. Bunker, Registration No. 29,365; William H. Nieman, Registration No. 30,201; John C. Lambertsen, Registration No. 29,400; Lowell Anderson, Registration No. 30,990; Arthur S. Rose, Registration No. 28,038; James F. Lesniak, Registration No. 25,240; Ned A. Israelsen, Registration No. 29,655; Drew S. Hamilton, Registration No. 29,801; Jerry T. Sewell, Registration No. 31,567; John B. Sganga, Jr., Registration No. 33,043; William C. Rooklidge, Registration No. 31,791; Joseph R. Re, Registration No. 31,921; John M. Carson, Registration No. 34,303; Andrew H. Simpson, Registration No. 31,469; Daniel E. Altman, Registration No. 34,115; Vito A. Canuso, Registration No. P-35, 471; William H. Shreve, Registration No. P-35, 678; Stephen C. Jensen, Registration No. P-35, 556; Anita M. Kirkpatrick, Registration No. 32,617; and Robert R. Devéza, Registration No. 33,230.

I hereby declare that all statements made herein of my own knowledge are true and that all statements made on information and belief are believed to be true; and further that these statements were made with the knowledge that willful false statements and the like so made are punishable by fine or imprisonment, or both, under Section 1001 of Title 18 of the United States Code and that such willful, false statement may jeopardize the validity of the application or any patent issued thereon.

1-00

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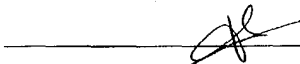
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<120> PROMOTER OF THIOREDOXINE TaTrxh2 IN  
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Trp Trp Leu Thr Ser Leu His His Gly Val  
45 50

